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Quantifying variation in female internal genitalia: no evidence for plasticity in response to sexual conflict risk in a seed beetle

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1 Abstract

Sexually antagonistic coevolution can drive the evolution of male traits that harm females, 2 3 and female resistance to those traits. While males have been found to vary their harmfulness to females in response to social cues, plasticity in female resistance traits remains to be 4 examined. Here we ask whether female seed beetles Callosobruchus maculatus are capable 5 6 of adjusting their resistance to male harm in response to the social environment. Among seed 7 beetles, male genital spines harm females during copulation and females might resist male harm via thickening of the reproductive tract walls. We develop a novel Micro-CT imaging 8 9 technique to quantify female reproductive tract thickness in 3-dimensional space, and compared the reproductive tracts of females from populations that had evolved under high 10 and low levels of sexual conflict, and for females reared under a social environment that 11 predicted either high or low levels of sexual conflict. We find little evidence to suggest that 12 females can adjust the thickness of their reproductive tracts in response to the social 13 environment. Neither did evolutionary history affect reproductive tract thickness. 14 Nevertheless, our novel methodology was capable of quantifying fine-scale differences in the 15 internal reproductive tracts of individual females, and will allow future investigations into the 16 internal organs of insects and other animals. 17

- 18 Key Words: female genital evolution, sexual conflict, sexually antagonistic coevolution,
- 19 phenotypic plasticity, experimental evolution, sex ratio

22 **1. Introduction**

Evolutionary conflict between the sexes can arise over the expression of traits that improve 23 the fitness of one sex but are detrimental to the fitness of their sexual partners [1,2]. Sexual 24 selection favours traits in males that confer greater fertilisation success regardless of the 25 outcomes for females, while antagonistic selection on females can favour morphological and 26 27 behavioural traits that function to resist harm induced by males [3,4]. Harmful traits in males are diverse in form and include: male harassment during courtship and mating [5,6], 28 infanticide [1,2], and toxic ejaculates [6,7]. One conspicuous form of sexual conflict is the 29 damage imposed to the female's reproductive tract by male genitalia during copulation [8]. 30 Genital damage during copulation is found across a variety of taxa, particularly within 31 arthropod lineages [9–14] and is considered to be a by-product of male traits enhancing 32 fertilisation success, rather than harm directly benefitting males [15–18]. 33

The evolution of harmful traits in one sex is expected to generate selection on the 34 opposite sex favouring the evolution of traits to resist harm, initiating sexually antagonistic 35 coevolution between the sexes [14,19–22]. While comparative evidence suggests that 36 coevolution of harmful male traits and female resistance traits is widespread [22–25], 37 whether these traits can respond plastically to an individual's environment has received less 38 attention. Males of a wide range of taxa have been found to adjust ejaculate size, composition 39 40 and quality in response to exposure to rival males [26-31], and recent work suggests that male genital morphology can be adjusted in response to the competitive environment 41 experienced during sexual development [32,33]. Thus, males exhibit phenotypic plasticity in 42 sexual traits that can be costly for females. Firman and Simmons [34] found that female 43 house mice exposed to greater levels of sperm competition risk produced ova with lower 44 fertilizability, presumably to mitigate against the threat of polyspermy when males compete 45 for fertilizations. This finding suggests that females too may be capable of responding to the 46

Page 6 of 28

socio-sexual environment, and reduce the costs associated with male competition. Using a
model species in which sexual conflict has been widely documented, we test the hypothesis
that sexual conflict favours the evolution of phenotypic plasticity in female resistance traits in
response to the immediate risk of sexual conflict.

Seed beetles (Callosobruchus maculatus) are a widely used model species for sexual 51 52 conflict studies [35]. Research has focused on the role and outcomes of male harmfulness on the persistence of polyandry, the effect of sperm competition on male ejaculate investment, 53 the effects of kinship on male harmfulness, and a wide array of other topics [16,18,36–38]. 54 The aedeagus (intromittent organ) is covered in sclerotized spines that perforate the female 55 reproductive tract during copulation, inflicting significant scarring [9,39]. The degree of 56 reproductive tract damage negatively impacts both female longevity and female reproductive 57 success [6] but promotes male fertilisation success [16]. Reproductive tract damage facilitates 58 the transport of accessory seminal compounds into the female bloodstream, which improve 59 male competitive fertilisation success [40–42]. Therefore, the harm to females in this species 60 seems to be a side-effect of selection for competitive male fertilization success [18,42]. The 61 thickness of the female reproductive tract wall appears to have evolved under sexually 62 antagonistic coevolution to resist the harmful effects of male genital spines. Thus, across 63 populations [22] and species [43], female reproductive tract volume has been found to be 64 positively correlated with male penile spine length, providing one of the few empirical 65 examples of sexually antagonistic coevolution [22]. Moreover, male C. maculatus appear to 66 adjust their copulatory behaviour [44,45] and amount of harm imposed on females in 67 response to their social environment [37,46, but see 47]. 68

Here, we investigate whether female *C. maculatus* exposed to greater risk of sexual
conflict can respond by adjusting the thickness of the reproductive tract to minimise
anticipated male harm. We used a manipulation of the social environment to vary the

immediate risk of sexual conflict: we had two treatments within which we manipulated both 72 larval density, and the adult sex-ratio to simulate either a high or low sexual conflict risk. 73 Additionally, we employed an experimental evolution design to test whether the thickness of 74 the female reproductive tract diverged between populations of beetles evolving under a male-75 or female-biased sex-ratio. Assessment of reproductive tract morphology in this species has 76 previously focused on either the thickness of the tract estimated from a small number of 77 histological sections [43], or the total volume of tissue across the entire tract via 3-78 dimensional analysis of Micro-CT images [22]. Although tissue volume may capture large-79 80 scale changes in female investment, it does not capture fine-scale changes in tract morphology. Here, we developed a novel technique to measure variation in the thickness of 81 the female reproductive tract at different locations along its length using Micro-CT data. We 82 predicted that: 1) females in populations evolving under a male-biased sex-ratio will have 83 84 thicker reproductive tract walls than those evolving under a female-biased sex ratio as a result of sexually antagonistic coevolution; and 2) females exposed to high-density larval 85 environments and a male-biased social environment during development will develop thicker 86 reproductive tract walls compared to females from low density larval environments exposed 87 to a female-biased social environment, in anticipation of an increased risk of sexual conflict. 88

89 **2. Methods**

90 *(a) Study population*

91 The stock population of *C. maculatus* used for this study was derived originally from a
92 population held by the CSIRO in 2005, which was itself founded by individuals found as
93 agricultural pests. Both stock and experimental populations were maintained at 30°C under a

94 12:12 hour day/night regime for the duration of the experiment [48]. For further information
95 regarding the stock population see [45].

96 *(b) Experimental evolution*

Experimental evolution lines were produced and maintained as described in McNamara et al. 97 [18]. In brief, individuals from the stock population were used to create six experimental 98 evolution lines. These lines were randomly assigned to one of two treatments, with either a 99 100 male- or female-biased sex ratio. Each generation consisted of 120 individuals, with an 80:40 male to female ratio for male-biased lines, and vice versa for female-biased lines. To control 101 for potential differences in larval competition between treatments, female-biased populations 102 103 received 200g of mung beans (Vigna radiata) for oviposition, while male-biased populations received 100g. Each new generation was created by isolating 300 beans per population within 104 ventilated 1.5mL Eppendorf tubes. After the required adults had emerged, sex-biased 105 populations were again formed. This procedure was continued for 47 generations, after which 106 populations were placed under common garden conditions for two generations with sex-ratio 107 108 parity to remove the potential for any non-genetic parental effects. Following the second generation of common-garden breeding, beans were placed within ventilated Eppendorf 109 tubes. 110

111 *(c) Social manipulation*

Isolated beans were assigned to one of two social manipulations designed to alter an individual's perception of future sexual conflict, either high-risk or low-risk. Evidence suggests that seed beetle larvae are able to determine population density before emergence from their beans via vibrations [49,50]. For this experiment we elected to use mung beans

(Vigna radiata) as the larval host species to increase surface area compared to their larger, 116 traditional host species (Vigna unguiculata), and thereby improve the transmission of 117 vibrational cues to focal individuals. Therefore, in the high-risk treatment, five infested 118 beans, each containing two larvae (infestation density can be assessed as eggs remain visible 119 on the surface of the bean), were placed within an Eppendorf tube (resulting in ten larvae 120 maximally per tube). Eppendorf tubes were checked daily for beetle emergence. Females that 121 122 emerged synchronously with a male were discarded from the experiment to ensure focal females had standardised pre-mating social exposure and remained unmated. Females that 123 124 emerged alone were placed within the lid of a 1.5mL Eppendorf tube, separated from four stock males and two stock females within the body of the tube via cotton mesh. The mesh 125 allowed focal females to detect the presence of individuals but prevented them from 126 copulating with the males. Previous studies have shown that this methodology elicits 127 phenotypic responses to the social environment in both sexes of C. maculatus [44,51]. Thus, 128 high-risk females experienced high larval density and a male-biased sex ratio. For the low-129 risk treatment, a single infested bean, containing a single larva, was placed within an 130 Eppendorf tube containing four un-infested beans. Following emergence, females were 131 placed within an Eppendorf tube separated from two stock males and four stock females. 132 Thus, low-risk females experienced a low larval density and a female-biased sex ratio prior to 133 mating. In the low-risk social treatment, we aimed to provide cues that were representative of 134 naturalistic conditions. Given the high densities experienced by C. maculatus when infesting 135 food stores, we therefore exposed females to a small number of males rather than no males. 136 For both treatments, females were removed after 24 hours and then allowed to mate once 137 with a stock male. Females were isolated for a further 24 hours post-copulation, and then 138 euthanized by freezing. Although we might expect adjustments in the reproductive tract to 139 most likely occur during larval and pupal development, we also included the adult 140

manipulation, because post-emergence sexual maturation is common in many insects, and
this species has been found to respond to manipulations of sexual conflict at the adult stage
[44,51].

144

145 *(d) Micro CT-scanning and tomographic reconstruction*

A total of five females per population-by-social-treatment combination were selected for
Micro-CT scanning. Therefore, a total of 60 individuals were scanned for the purposes of this
study. Sample tissue staining and scanning procedures followed those outlined by Dougherty
et al. [22], with the exception that we used formalin for tissue fixation rather than
paraformaldehyde (see detailed pre-scan methodology in the online supplementary material).

151 Samples were scanned using a ZEISS Xradia Versa 520 X-ray microscope housed at the University of Western Australia Centre for Microscopy, Characterisation and Analysis. 152 153 Samples were suspended in 100% ethanol and mounted in heat-sealed pipette tips with waxcovered tops in groups of 3-5 for scanning. Abdomens were arranged vertically and scanned 154 sequentially from the highest abdomen to the lowest. Source voltage and power of initial 155 scans (n = 11) was set at 40kV and 3W. However, source stability was compromised for an 156 extended period of time, so the remaining scans were conducted at 60kV and 5W (n = 43) to 157 ensure that scan quality remained stable (details of the scanning procedures can be found in 158 the online supporting material). A total of six samples could not be analysed due to low scan 159 quality, leaving a sample size of 54 for image analysis. 160

161

162 *(e) Image analysis*

Images were analysed blind with regard to both the population of origin and social treatment. 163 The analyses were performed using a combination of three custom-written FIJI [52–54] 164 scripts and Amira 6.2 (Thermo-Fisher Scientific, U.S.A.). We first selected a consistent 165 region of interest within each reproductive tract, which was marked by the entrance of the 166 spermathecal duct into the reproductive tract at one end (Figure 1a) and the first occurrence 167 of bursal teeth on the other (Figure 1b) This region was chosen because it sustains the 168 greatest damage during copulation [22]. Within this region of interest, we computed the 3D 169 thickness of the dorsal and ventral reproductive tract walls based on the local thickness 170 171 definition proposed by Dougherty and Kunzelmann [55] (see online Supplementary material for detailed methodology). According to this definition the thickness at any point within an 172 object, in our case the tract walls, is the diameter of the largest sphere that fits inside the 173 object and at the same time contains the point (Figure 1c). Further, we differentiated 174 investment in the upper and lower reproductive tract by placing a horizontal plane running 175 through the lumen's centroid, which allowed us to reliably define the upper and lower regions 176 of tracts among all sampled individuals (Figure 1d). 177

178

179 *(f) Statistical Analyses*

All statistical analyses were conducted in R (v 3.5.3) [56]. Normality of data was confirmed
with Shapiro-Wilk's tests. Further, Bartlett's test for homogeneity of variance was found to
be non-significant in all cases.

Pearson's correlation tests using the 'ppcor' R package [57] found significant correlations between the mean, minimum and maximum thickness values from the upper and lower reproductive tract. Repeatability analyses were conducted for all measurements by extracting measures of 9 individuals on each of three separate occasions and analysing the data using

Page 12 of 28

the R package 'rptR' [58]. We found significant repeatability (R > 0.6) in mean and 187 maximum tract thickness but not in minimum tract thickness (see Table S2 in the online 188 Supplementary Material). We therefore conducted principal components analysis using the 189 'FactoMineR' R package, excluding upper and lower minimum thickness [59]. One principal 190 component (PC1) had an eigenvalue greater than 1 and was extracted for further analysis. 191 PC1 was used as the dependent variable using a Gaussian linear mixed model that included 192 193 female weight as a covariate. Following Arnqvist [60], replicate population was included as a random factor within which an interaction between female weight and social treatment was 194 195 fitted as a random slope. The random slope allows for the interaction to vary across the differing populations [60]. The source voltage was also included in this model as a random 196 factor to control for any variance in trait estimates that might have arisen from the use of 197 different voltage settings during scanning. The significance of our treatments, random factors 198 199 and interaction effects were tested using Kenward-Roger F-tests using the R package 'pbkrtest'[61]. Non-significant interactions that did not improve model fit were subsequently 200 removed from final models. Effect sizes were estimated as the standardised Pearson's 201 correlation coefficients (Table 3) using the 'effectsize' package in R. Estimated marginal 202 means for each level of the two treatments were produced via the package 'emmeans'. The 203 data were explored for outliers using robust kernel-based outlier factor algorithms within the 204 205 'OutlierDetection' R package (k=3, bootstraps= 50,000). No outliers were identified.

206 **3. Results**

The first principal component explained 72% of the variance in reproductive tract thickness (Table 1) and was loaded equally by the mean and maximum thickness from both the upper and lower reproductive tract. There was no significant impact of evolutionary history or

210	social treatment on female tract (Table 2). All interaction combinations between evolutionary
211	history, social treatment and weight were found to be non-significant and were dropped from
212	the final model.

- 213
- 214

215 4. Discussion

We developed a novel Micro-CT imaging method to measure the thickness of the 216 reproductive tract walls of female C. maculatus seed beetles, from populations that had 217 evolved under a male- or female-biased population sex ratio for 47 generations, and which 218 were subsequently exposed to a social environment which conveyed either a high or low-risk 219 220 of sexual conflict. Previous studies of this species have found that males can adjust their harmfulness to females in response to their social environment [43–45, but see: 56]. There 221 222 was no effect of evolutionary history on the overall thickness of the female reproductive tract, 223 nor was there any effect of the social environment. Our data therefore suggests that female reproductive tracts may not respond to variation in the risk of sexual conflict. 224

Previous studies using *C. maculatus* have similarly failed to find evolutionary 225 responses in females to experimental manipulations of sexual conflict. Gay et al. [57] showed 226 that after 90 generations of enforced monogamy, the re-introduction of polyandry over 30 227 generations resulted in the evolution of harmful males but not resistant females. Similarly, 228 after 8 generations of enforced monogamy it was found that the elaboration of male penile 229 230 spines decreased, as might be predicted for a costly trait that functions in the context of competitive reproductive success. However, the morphology of the teeth found within the 231 female genital tract failed to exhibit a correlated response to enforced monogamy [64]. Our 232 results are also consistent with recent findings of McNamara et al. [18] who utilised the same 233

experimental evolution lines used in the current study. McNamara et al. [18] found that 234 although males evolving under a male-biased sex ratio evolved to be more harmful, females 235 from populations evolving under a male-biased sex ratio experienced comparable 236 reproductive tract scarring to females from populations evolving under a female-biased sex 237 ratio. We found no evidence that sexual conflict intensity results in an evolutionary 238 divergence in female reproductive tract thickness. Collectively, these findings suggest that 239 240 either females fail to coevolve as readily as males, or that female reproductive tract coevolution is more difficult to detect [18,63,64]. 241

The findings of these experimental evolution studies are in contrast to comparative 242 studies that have found evidence for sexually antagonistic coevolution between male 243 harmfulness and female resistance to harm among C. maculatus populations and among 244 *Callosobruchus* species more widely [22,43]. Further, the coevolution of resistance traits and 245 male persistence has been identified in comparative studies of other arthropod taxa such as 246 water striders (Heteroptera: Gerridae) [65] and diving beetles (Coleoptera: Dytiscidae) [66]. 247 The apparent inability of female seed beetle reproductive tracts to respond evolutionarily to 248 relatively short-term manipulations in sexual conflict might be attributable to the difficulty in 249 detecting sexually antagonistic coevolution at a particular point in time [2,24,67]. This 250 suggestion could explain why females seemingly did not respond to our selection treatments, 251 252 as it is clear that divergence in male harmfulness was present in these populations 15 generations earlier [18]. We are unable to say whether females from populations experiencing 253 differing levels of sexual conflict might have developed alternative methods of resistance, 254 such as higher immunocompetence [68]. Previous studies utilising the same experimental 255 evolution lines have shown that individuals derived from the male-biased lines exhibit 256 reduced immune function [51]. These results are indicative of a resource trade-off between 257 immune function and reproductive investment fuelled by the costs of high intensity sexual 258

conflict. Although the method by which Gay et al. [57] and McNamara et al. [18] altered 259 sexual conflict intensity differed, both studies found that females from populations that 260 experienced higher conflict were better able to counter-act the negative impact of mating on 261 their fitness. However, it is clear that the method by which female *C. maculatus* accomplish 262 this is not via genital morphology, female kicking behaviour, or through improved immunity 263 [18,51]. Finally, it may be that inbreeding depression was responsible for impeding 264 265 divergence among our lines. This seems unlikely however, because a hallmark of inbreeding is reduced fitness, which would have been observed as reduced fecundity for females from 266 267 the female-biased lines. A previous study using the same experimental evolution lines [18] showed no evidence that females from female-biased lines experienced reduce fitness 268 compared to those from male-biased lines. 269

270 We found that the social environment had no impact on female reproductive tract thickness. This is the only study to have investigated whether females are able to plastically 271 respond to sexual conflict risk by altering reproductive tissue dimensions. There are several 272 reasons why females might have failed to respond to our manipulation of their social 273 environment. First, it is possible that females were unable to adjust the reproductive tract in 274 response to our environmental cues. Females were given a proxy for sexual conflict risk via 275 manipulations of larval density during development, but this cue may have been insufficient 276 277 to provoke a plastic response. Second, the larval and pupal stages are critical to the development of insect reproductive organs [69]. This is demonstrated by the long-lasting 278 impacts of larval food availability on adult reproductive output in both female and male 279 insects [70,71]. Females were exposed to direct signals of increased sexual conflict risk (via 280 sex-ratio) in their adult stage, post-pupation, but they may be unable to make plastic 281 adjustments after adult emergence. Third, it is also possible that our measures of tract 282 thickness do not capture important qualitative variation. For example, it may be that females 283

can also plastically adjust the elasticity of the reproductive tract through the incorporation of 284 resilin, a compound shown to improve tolerance to cuticle perforation in bed bugs (Cimex 285 *lectularius*)[72]. Finally, it may be that females plastically responded to our social 286 manipulation, but due to the logistical limitations placed on us with respect to the numbers of 287 females we could scan, we were unable to capture any differences among them. However, 288 given that our repeatability analysis revealed significant variation among females in 289 290 reproductive tract thickness, we can be certain that we are able to detect variation in tract thickness using this novel technique. If there were an effect of the social environment on the 291 292 reproductive tract thickness, the effect size estimated in our analysis suggest that it is small and would require a large sample size in order to detect significance. 293

294

295 Although our measure of female resistance traits is limited to a quantitative measure of reproductive tract thickness, it nonetheless offers a significant advance over previous 296 297 studies. Previous studies utilising 3-dimensional analysis of Micro-CT scans have measured total reproductive tract volume, which is effective at controlling for tract shape and size 298 effects, but cannot identify fine-scale changes in morphology within the reproductive tract 299 [22]. Our current method overcomes this challenge by allowing us to identify variation in 300 thickness in the upper and lower regions of the reproductive tract in 3-dimensional space. 301 302 Overall, our novel method for measuring reproductive tract thickness shows promise in its ability to detect fine-scale differences in internal structures of the female reproductive tract, 303 and promises to be a valuable tool in the long-awaited study of female genital morphology 304 across numerous species. 305

In conclusion, we provide no evidence for plastic adjustments of reproductive tractthickness, a trait known to have coevolved with male-imposed genital damage among

populations and species of Callosobruchus. Moreover, we found that populations that had 308 evolved under intense sexual conflict failed to diverge in female genital morphology. The 309 coevolution of male and female genital traits among populations suggests ample genetic 310 variation in tract morphology may exist, however among-population genetic variance does 311 not necessarily reflect within-population variation [73]. Therefore, the lack of divergence in 312 tract morphology across our evolution lines may reflect a lack of within-population genetic 313 314 variation for this trait. Studies of sexual selection acting on female genitalia typically lag behind those focussed on male traits [67,74]. Investigating female traits and their responses to 315 316 male harmfulness can broaden our understanding of sexual selection and sexual conflict. Further research needs to focus on the female perspective if we are to quantify the 317 pervasiveness and intensity of female responses to sexual conflict. The barriers to such 318 research are slowly dissipating with the advent of innovative new technologies, such as 319 Micro-CT scanning, that allow more effective measurement of female traits. As technologies 320 become more accessible and cheaper to employ, increased sample sizes will be possible so 321 that future studies have the power to detect variation in these minute structures. We believe 322 that the present study provides a viable methodological approach for further investigations 323 into plastic adjustments in female reproductive tracts, which is flexible enough to identify 324 small-scale variation. 325

326

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337	Chara	cterisation and Analysis (CMCA), The University of Western Australia.	
338			
339		References	
340	1.	Parker GA. 2006 Sexual conflict over mating and fertilization: An overview. Philos. Trans. R.	
341		Soc. B Biol. Sci. 361, 235–259. (doi:10.1098/rstb.2005.1785)	
342	2.	Arnqvist G, Rowe L. 2005 Sexual Conflict. Princeton, N.J.: Princeton University Press.	
343	3.	Stockley P. 1997 Sexual conflict resulting from adaptations to sperm competition. Trends	
344		Ecol. Evol. 12, 154–159. (doi:https://doi.org/10.1016/S0169-5347(97)01000-8)	
345	4.	Edvardsson M, Tregenza T. 2005 Why do male Callosobruchus maculatus harm their mates?	
346		Behav. Ecol. 16, 788–793. (doi:10.1093/beheco/ari055)	
347	5.	Rankin DJ, Dieckmann U, Kokko H. 2011 Sexual Conflict and the Tragedy of the Commons.	
348		Am. Nat. 177, 780-791. (doi:10.1086/659947)	
349	6.	den Hollander M, Gwynne DT. 2009 Female fitness consequences of male harassment and	
350		copulation in seed beetles, Callosobruchus maculatus. Anim. Behav. 78, 1061-1070.	
351		(doi:10.1016/j.anbehav.2009.06.036)	
352	7.	Chapman T, Liddle LF, Kalb JM, Wolfner MF, Partridge L. 1995 Cost of mating in	
353		Drosophila melanogaster females is mediated by male accessory gland products. <i>Nature</i> . 373,	
354		241-244. (doi:10.1038/373241a0)	

Page 19 of 28

355	8.	Lange R, Reinhardt K, Michiels NK, Anthes N. 2013 Functions, diversity, and evolution of
356		traumatic mating. Biol. Rev. 88, 585-601. (doi:10.1111/brv.12018)
357	9.	Crudgington HS, Siva-Jothy MT. 2000 Genital damage, kicking and early death. Nature 407,
358		855-856. (doi:10.1038/35038154)
359	10.	Blanckenhorn WU, Hosken DJ, Martin OY, Reim C, Teuschl Y, Ward PI. 2002 The costs of
360		copulating in the dung fly Sepsis cynipsea. Behav. Ecol. 13, 353–358.
361		(doi:10.1093/beheco/13.3.353)
362	11.	Kamimura Y. 2012 Correlated evolutionary changes in Drosophila female genitalia reduce the
363		possible infection risk caused by male copulatory wounding. Behav. Ecol. Sociobiol. 66,
364		1107–1114. (doi:10.1007/s00265-012-1361-0)
365	12.	Kamimura Y. 2010 Copulation anatomy of Drosophila melanogaster (Diptera: Drosophilidae):
366		wound-making organs and their possible roles. Zoomorphology 129, 163–174.
367		(doi:10.1007/s00435-010-0109-5)
368	13.	Tatarnic NJ, Cassis G, Siva-Jothy MT. 2014 Traumatic Insemination in Terrestrial Arthropods.
369		Annu. Rev. Entomol. 59, 245–261. (doi:10.1146/annurev-ento-011613-162111)
370	14.	Tatarnic NJ, Cassis G. 2010 Sexual coevolution in the traumatically inseminating plant bug
371		genus Coridromius. J. Evol. Biol. 23, 1321–1326. (doi:10.1111/j.1420-9101.2010.01991.x)
372	15.	Morrow EH, Arnqvist G, Pitnick S. 2003 Adaptation versus pleiotropy: Why do males harm
373		their mates? Behav. Ecol. 14, 802-806. (doi:10.1093/beheco/arg073)
374	16.	Hotzy C, Arnqvist G. 2009 Sperm Competition Favors Harmful Males in Seed Beetles. Curr.
375		<i>Biol.</i> 19 , 404–407. (doi:10.1016/j.cub.2009.01.045)
376	17.	Grieshop K, Polak M. 2014 Evaluating the post-copulatory sexual selection hypothesis for
377		genital evolution reveals evidence for pleiotropic harm exerted by the male genital spines of
378		Drosophila ananassae. J. Evol. Biol. 27, 2676–2686. (doi:10.1111/jeb.12524)

18.

McNamara KB, Sloan NS, Kershaw SE, van Lieshout E, Simmons LW. 2020 Males evolve to

380		be more harmful under increased sexual conflict intensity in a seed beetle. Behav. Ecol.
381		(doi:10.1093/beheco/arz186)
382	10	Arnavist G. Rowe I 1995 Sexual conflict and arms races between the sexes: a mornhological
302	19.	Aniquist G, Rowe E. 1999 Sexual connect and arms faces between the sexes. a morphological
383		adaptation for control of mating in a female insect. Proc. R. Soc. London. Ser. B Biol. Sci. 261,
384		123–127. (doi:10.1098/rspb.1995.0126)
385	20.	Rice WR. 1996 Sexually antagonistic male adaptation triggered by experimental arrest of
386		female evolution. Nature 381, 232.
387	21.	Arnqvist G, Rowe L. 2002 Antagonistic coevolution between the sexes in a group of insects.
388		<i>Nature</i> 415 , 787–789.
389	22.	Dougherty LR, van Lieshout E, McNamara KB, Moschilla JA, Arnqvist G, Simmons LW.
390		2017 Sexual conflict and correlated evolution between male persistence and female resistance
391		traits in the seed beetle Callosobruchus maculatus. <i>Proc. R. Soc. B Biol. Sci.</i> 284, 20170132.
392		(doi:10.1098/rspb.2017.0132)
393	23.	Brennan PLR, Prum RO, McCracken KG, Sorenson MD, Wilson RE, Birkhead TR. 2007
394		Coevolution of Male and Female Genital Morphology in Waterfowl. PLoS One 2, e418.
395	24.	Perry JC, Rowe L. 2012 Sexual conflict and antagonistic coevolution across water strider
396		populations. Evolution (N. Y). 66, 544–557. (doi:10.1111/j.1558-5646.2011.01464.x)
397	25.	Hopwood PE, Head ML, Jordan EJ, Carter MJ, Davey E, Moore AJ, Royle NJ. 2016 Selection
398		on an antagonistic behavioral trait can drive rapid genital coevolution in the burying beetle.
399		Nicrophorus vespilloides. <i>Evolution</i> 70, 1180–1188. (doi:10.1111/evo.12938)
400	26.	DelBarco-Trillo J. 2011 Adjustment of sperm allocation under high risk of sperm competition
401		across taxa: a meta-analysis. J. Evol. Biol. 24, 1706–1714. (doi:10.1111/j.1420-
402		9101.2011.02293.x)
403	27.	Thomas ML, Simmons LW. 2007 Male Crickets Adjust the Viability of Their Sperm in

Page 21 of 28

404		Response to Female Mating Status. Am. Nat. 170, 190–195. (doi:10.1086/519404)
405	28.	Simmons LW, Denholm A, Jackson C, Levy E, Madon E. 2007 Male crickets adjust ejaculate
406		quality with both risk and intensity of sperm competition. Biol. Lett. 3, 520-522.
407		(doi:10.1098/rsbl.2007.0328)
408	29.	Gage MJG, Baker RR. 1991 Ejaculate size varies with socio-sexual situation in an insect. Ecol.
409		<i>Entomol.</i> 16 , 331–337. (doi:10.1111/j.1365-2311.1991.tb00224.x)
410	30.	Evans JP, Pierotti M, Pilastro A. 2003 Male mating behavior and ejaculate expenditure under
411		sperm competition risk in the eastern mosquitofish. Behav. Ecol. 14, 268-273.
412	31.	Pilastro A, Scaggiante M, Rasotto MB. 2002 Individual adjustment of sperm expenditure
413		accords with sperm competition theory. Proc. Natl. Acad. Sci. 99, 9913 LP - 9915.
414		(doi:10.1073/pnas.152133499)
415	32.	Brennan PLR, Gereg I, Goodman M, Feng D, Prum RO. 2017 Evidence of phenotypic
416		plasticity of penis morphology and delayed reproductive maturation in response to male
417		competition in waterfowl. Auk 134, 882–893. (doi:10.1642/AUK-17-114.1)
418	33.	André GI, Firman RC, Simmons LW. 2018 Phenotypic plasticity in genitalia: baculum shape
419		responds to sperm competition risk in house mice. Proc. R. Soc. B Biol. Sci. 285, 20181086.
420		(doi:10.1098/rspb.2018.1086)
421	34.	Firman RC, Simmons LW. 2013 Sperm competition risk generates phenotypic plasticity in
422		ovum fertilizability. Proc. R. Soc. B Biol. Sci. 280, 20132097. (doi:10.1098/rspb.2013.2097)
423	35.	Zuk M, García-González F, Herberstein ME, Simmons LW. 2014 Model systems, taxonomic
424		bias, and sexual selection: Beyond drosophila. Annu. Rev. Entomol. 59, 321-338.
425		(doi:10.1146/annurev-ento-011613-162014)
426	36.	Wilson CJ, Tomkins JL. 2015 Female Callosobruchus maculatus can maximize long-term
427		fitness through polyandry. Behav. Ecol. 26, 502-509. (doi:10.1093/beheco/aru218)

- 428 37. Lymbery SJ, Simmons LW. 2017 Males harm females less when competing with familiar
 429 relatives. *Proc. R. Soc. B Biol. Sci.* 284, 20171984. (doi:10.1098/rspb.2017.1984)
- 430 38. Rönn JL, Katvala M, Arnqvist G. 2006 The costs of mating and egg production in
- 431 Callosobruchus seed beetles. *Anim. Behav.* **72**, 335–342.
- 432 (doi:https://doi.org/10.1016/j.anbehav.2005.10.024)
- 433 39. Dougherty LR, Simmons LW. 2017 X-ray micro-CT scanning reveals temporal separation of
- 434 male harm and female kicking during traumatic mating in seed beetles. *Proc. R. Soc. B Biol.*

435 *Sci.* **284**. (doi:10.1098/rspb.2017.0550)

- 436 40. Yamane T, Goenaga J, Rönn JL, Arnqvist G. 2015 Male seminal fluid substances affect sperm
 437 competition success and female reproductive behavior in a seed beetle. *PLoS One* 10, 1–14.
- 438 (doi:10.1371/journal.pone.0123770)
- 439 41. Rönn JL, Hotzy C. 2012 Do longer genital spines in male seed beetles function as better
 440 anchors during mating? *Anim. Behav.* 83, 75–79. (doi:10.1016/j.anbehav.2011.10.007)
- 441 42. Hotzy C, Polak M, Rönn JL, Arnqvist G. 2012 Phenotypic Engineering Unveils the Function
 442 of Genital Morphology. *Curr. Biol.* 22, 2258–2261.
- 443 (doi:https://doi.org/10.1016/j.cub.2012.10.009)
- 444 43. Ronn J, Katvala M, Arnqvist G. 2007 Coevolution between harmful male genitalia and female
 445 resistance in seed beetles. *Proc. Natl. Acad. Sci.* 104, 10921–10925.
- 446 (doi:10.1073/pnas.0701170104)
- 447 44. Wilson CJ, Tomkins JL. 2014 Countering counteradaptations: Males hijack control of female
 448 kicking behavior. *Behav. Ecol.* 25, 470–476. (doi:10.1093/beheco/aru022)
- 449 45. Wilson CJ, Buzatto BA, Robinson SP, Tomkins JL. 2014 Sociosexual environment influences
- 450 patterns of ejaculate transfer and female kicking in Callosobruchus maculatus. *Anim. Behav.*
- 451 94, 37–43. (doi:https://doi.org/10.1016/j.anbehav.2014.05.014)
- 452 46. Lymbery SJ, Tomkins JL, Simmons LW. 2019 Male responses to sperm competition when

453 rivals vary in number and familiarity. *Proc. R. Soc. B Biol. Sci.* 286.

- 454 (doi:10.1098/rspb.2018.2589)
- 47. Berg EC, Lind MI, Monahan S, Bricout S, Maklakov AA. 2019 Kin but less than kind: withingroup male relatedness does not increase female fitness in seed beetles. *Proc. R. Soc. B Biol.*
- 457 *Sci.* **286**, 20191664. (doi:10.1098/rspb.2019.1664)
- 458 48. McNamara KB, Robinson SP, Rosa ME, Sloan NS, van Lieshout E, Simmons LW. 2016
- 459 Male-biased sex ratio does not promote increased sperm competitiveness in the seed beetle,
 460 Callosobruchus maculatus. *Sci. Rep.* 6, 28153.
- 461 49. Utida S. 1972 Density dependent polymorphism in the adult of Callosobruchus maculatus
 462 (Coleoptera, Bruchidae). *J. Stored Prod. Res.* 8, 111–125. (doi:https://doi.org/10.1016/0022-
- 463 474X(72)90028-8)
- 464 50. Thanthianga C, Mitchell R. 1987 Vibrations mediate prudent resource exploitation by
 465 competing larvae of the bruchid bean weevil Callosobruchus maculatus. *Entomol. Exp. Appl.*466 44, 15–21. (doi:10.1111/j.1570-7458.1987.tb02233.x)
- 467 51. van Lieshout E, McNamara KB, Simmons LW. 2014 Rapid Loss of Behavioral Plasticity and
- 468 Immunocompetence Under Intense Sexual Selection. *Evolution (N. Y).* 68, 2550–2558.
- 469 (doi:10.1111/evo.12422)
- 470 52. Schneider CA, Rasband WS, Eliceiri KW. 2012 NIH Image to ImageJ: 25 years of image
 471 analysis. *Nat. Methods*. (doi:10.1038/nmeth.2089)
- 472 53. Legland D, Arganda-Carreras I, Andrey P. 2016 MorphoLibJ: integrated library and plugins
- for mathematical morphology with ImageJ. *Bioinformatics* **32**, 3532–3534.
- 474 (doi:10.1093/bioinformatics/btw413)
- 475 54. Schindelin J *et al.* 2012 Fiji: an open-source platform for biological-image analysis. *Nat.*476 *Methods* 9, 676–682. (doi:10.1038/nmeth.2019)
- 477 55. Dougherty R, Kunzelmann K-H. 2007 Computing Local Thickness of 3D Structures with

478		ImageJ. Microsc. Microanal. 13, 1678–1679. (doi:DOI: 10.1017/S1431927607074430)
479	56.	R Development Core Team. 2016 R: A language and environment for statistical computing. R
480		Found. Stat. Comput. (doi:10.1017/CBO9781107415324.004)
481	57.	Kim S. 2015 ppcor: An R Package for a Fast Calculation to Semi-partial Correlation
482		Coefficients. Commun. Stat. Appl. methods 22, 665-674. (doi:10.5351/CSAM.2015.22.6.665)
483	58.	Stoffel MA, Nakagawa S, Schielzeth H. 2017 rptR: repeatability estimation and variance
484		decomposition by generalized linear mixed-effects models. Methods Ecol. Evol. 8, 1639–1644.
485		(doi:10.1111/2041-210X.12797)
486	59.	Lê S, Josse J, Husson F. 2008 FactoMineR: An R Package for Multivariate Analysis. J. Stat.
487		Software; Vol 1, Issue 1
488	60.	Arnqvist G. 2020 Mixed Models Offer No Freedom from Degrees of Freedom. Trends Ecol.
489		Evol. 35, 329–335. (doi:https://doi.org/10.1016/j.tree.2019.12.004)
490	61.	Halekoh U, Højsgaard S. 2014 A Kenward-Roger Approximation and Parametric Bootstrap
491		Methods for Tests in Linear Mixed Models The R Package pbkrtest. J. Stat. Software; Vol 1,
492		Issue 9 (doi:10.18637/jss.v059.i09)
493	62.	Le Page S, Sepil I, Flintham E, Pizzari T, Carazo P, Wigby S. 2017 Male relatedness and
494		familiarity are required to modulate male-induced harm to females in Drosophila. Proc. R.
495		Soc. B Biol. Sci. 284, 11–14. (doi:10.1098/rspb.2017.0441)
496	63.	Gay L, Hosken DJ, Eady PE, Vasudev R, Tregenza T. 2010 The evolution of harm-effect of
497		sexual conflicts and population size. Evolution (N. Y). 65, 725-737. (doi:10.1111/j.1558-
498		5646.2010.01181.x)
499	64.	Cayetano L, Maklakov AA, Brooks RC, Bonduriansky R. 2011 Evolution of male and female
500		genitalia following release from sexual selection. Evolution (N. Y). 65, 2171–2183.
501		(doi:10.1111/j.1558-5646.2011.01309.x)

502	65.	Arnqvist G, Rowe L. 2002 Correlated evolution of male and female morphologies in water
503		striders. Evolution (N. Y). 56, 936–947. (doi:10.1111/j.0014-3820.2002.tb01406.x)
504	66.	Bergsten J, Töyrä A, Nilsson AN. 2008 Intraspecific variation and intersexual correlation in
505		secondary sexual characters of three diving beetles (Coleoptera: Dytiscidae). Biol. J. Linn. Soc.
506		73 , 221–232. (doi:10.1111/j.1095-8312.2001.tb01359.x)
507	67.	Kokko H, Jennions MD. 2014 The relationship between sexual selection and sexual conflict.
508		Cold Spring Harb. Perspect. Biol. 6. (doi:10.1101/cshperspect.a017517)
509	68.	Hangartner S, Michalczyk Ł, Gage MJG, Martin OY. 2015 Experimental removal of sexual
510		selection leads to decreased investment in an immune component in female Tribolium
511		castaneum. Infect. Genet. Evol. 33, 212–218.
512		(doi:https://doi.org/10.1016/j.meegid.2015.05.005)
513	69.	Happ GM. 1992 Maturation of the Male Reproductive System and its Endocrine Regulation.
514		Annu. Rev. Entomol. 37, 303–320. (doi:10.1146/annurev.en.37.010192.001511)
515	70.	Engels S, Sauer KP. 2007 Energy beyond the pupal stage: Larval nutrition and its long-time
516		consequences for male mating performance in a scorpionfly. J. Insect Physiol. 53, 633-638.
517		(doi:https://doi.org/10.1016/j.jinsphys.2007.05.003)
518	71.	Bauerfeind SS, Fischer K. 2005 Effects of food stress and density in different life stages on
519		reproduction in a butterfly. Oikos 111, 514-524. (doi:https://doi.org/10.1111/j.0030-
520		1299.2005.13888.x)
521	72.	Michels J, Gorb SN, Reinhardt K. 2015 Reduction of female copulatory damage by resilin
522		represents evidence for tolerance in sexual conflict. J. R. Soc. Interface 12, 20141107.
523		(doi:10.1098/rsif.2014.1107)
524	73.	Hoffmann AA, Hallas RJ, Dean JA, Schiffer M. 2003 Low Potential for Climatic Stress
525		Adaptation in a Rainforest Drosophila Species. Science (80). 301, 100 LP – 102.
526		(doi:10.1126/science.1084296)

- 527 74. Sloan NS, Simmons LW. 2019 The evolution of female genitalia. J. Evol. Biol. 32, 882–899.
- **528** (doi:10.1111/jeb.13503)

- **Table 1** Fit and loadings for the first two principal components explaining variation in female
- 531 reproductive tract thickness.

	PC1	PC2
Eigenvalue	2.88	0.78
% Variance	71.91	19.91
Upper mean	0.930	0.25
Upper maximum	0.777	0.60
Lower mean	0.877	-0.36
Lower maximum	0.811	-0.47

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Table 2 Analysis of variance for fixed and random effects on a multivariate measure of

	-	F	df	р	Variance
	Evolutionary History	0.51	1, 3.79	0.52	
	Social Environment (SE)	1.18	1, 4.38	0.34	
	Body Weight	3.95	1, 3.61	0.12	
	Population replicate				4.31
	Source voltage				< 0.01
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535 female reproductive tract thickness (PC1) (N=54).

- **Table 3** Estimated marginal means and effect sizes (Pearson's r) with 95% confidence
- 546 intervals of all fixed effects on a multivariate measure of female reproductive tract thickness
- 547 (PC1) (N=54).

	Mean	95% CI	Effect size	95% CI
			(r)	
Evolutionary History (M-F)			-0.28	-0.76, 0.21
Female-bias	0.638	0.013, 1.26		
Male-bias	0.580	-0.17, 1.33		
Social Environment (L-H)			-0.43	-0.92, 0.05
High-risk	0.643	0.047, 1.24		
Low-risk	0.575	-0.15, 1.30		



Segmentation of the reproductive tract: (a) First appearance of the entrance of the spermathecal duct. (b) First appearance of the bursal teeth. Tract wall thickness: (c) 3D thickness heat map (d) Upper and lower regions of interest within the reproductive tract.

444x380mm (130 x 130 DPI)